

Patterned Flow Cell Technology

Fixed spacing of sequencing clusters with defined feature sizes contributes to increased data output, reduced costs, and faster run times.

Introduction

Leveraging the highly accurate and robust sequencing by synthesis (SBS) technology, Illumina sequencing platforms are the most successful and widely adopted next-generation sequencing (NGS) systems worldwide. Patterned flow cell technology significantly increases the throughput of Illumina NGS systems, drastically reducing the sequencing cost per sample. This price decrease enables data-intensive applications such as whole-genome sequencing for population-scale studies, including the \$1000 human genome, on the HiSeq X™ Ten and HiSeq X Five Systems.^{*} The HiSeq® 3000/HiSeq 4000 Systems[†] take advantage of this technology improvement to offer a cost-effective solution for a broad range of applications.

Higher-Density Clonal Clusters

Two innovative breakthroughs contribute to the boost in sequencing power experienced with patterned flow cell technology: 1) a distinct, ordered nanowell design and 2) a new exclusion amplification chemistry. The result is more efficient use of the flow cell surface area yielding a higher density of sequenceable clusters during cluster generation. Higher cluster density leads to more useable data per flow cell, driving down the cost per gigabase (Gb) of the sequencing run.

Impact of Cluster Generation

The basic Illumina NGS workflow includes 4 steps: 1) library preparation, 2) cluster generation, 3) sequencing, and 4) alignment and data analysis. Historically, during cluster generation, adapter-ligated library elements hybridized to complementary oligonucleotides on the surface of a flow cell. Each attached library fragment acted as a “seed” and, through a process called bridge amplification, was amplified to generate a clonal cluster containing thousands of identical fragments. After cluster generation was complete, the flow cell contained millions to billions of clusters on its surface.

Ideally, clusters are of similar size and spaced well apart from each other to achieve accurate resolution during imaging. In reality, DNA clusters are randomly distributed across the flow cell with many clusters in close proximity to neighboring clusters, especially if the sample is overloaded, making it difficult to discern individual clusters from each other. If a cluster cannot be accurately resolved from an adjacent cluster, the data from that cluster is considered unusable, reducing the amount of information generated during the run. Patterned flow cell technology overcomes these challenges with prearranged nanowells that optimize cluster spacing. Clusters can only form in the nanowells, making the flow cells less susceptible to

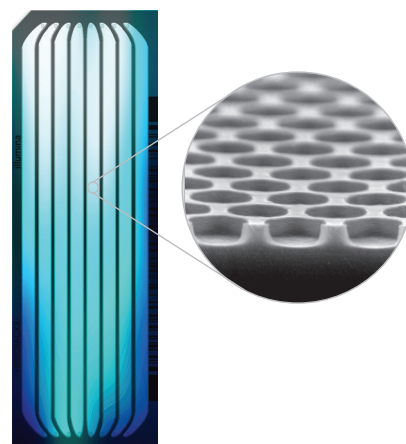


Figure 1: Patterned Flow Cell Design—Patterned flow cells contain billions of nanowells at fixed locations, providing even cluster spacing and uniform feature size to deliver extremely high cluster densities.

overloading. New patented chemistry allows simultaneous seeding and amplification during cluster generation to minimize the chances of multiple library fragments amplifying as a single cluster.

Prearranged Cluster Distribution on the Flow Cell

To make more effective use of the flow cell surface space, Illumina created the patterned flow cell with distinct nanowells for cluster generation (Figure 1). Patterned flow cells are produced using semiconductor manufacturing technology. Starting with a glass substrate, patterned nanowells are etched into the surface. Each nanowell contains DNA probes used to capture prepared DNA strands for amplification during cluster generation. The area between the nanowells is devoid of DNA probes, preventing the formation of clusters in the interstitial regions between wells.

This process ensures that DNA clusters only form within the nanowells, providing even, consistent spacing between adjacent clusters and allowing accurate resolution of clusters during imaging. The result is maximal use of the flow cell surface leading to overall higher clustering.

Exclusion Amplification

By enabling simultaneous seeding (landing of the DNA strand in the nanowell) and amplification, exclusion amplification promotes monoclonal cluster generation within the nanowells. This improvement allows the number of monoclonal clusters available for sequencing on a patterned flow cell to exceed the Poisson statistical limit, significantly increasing data output.

^{*} The HiSeq X Ten System is a suite of 10 individual HiSeq X instruments. The HiSeq X Five System is a suite of 5 individual HiSeq X instruments.

[†] The HiSeq 4000 System is a dual-flow cell instrument with higher throughput than the single-flow cell HiSeq 3000 System.

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