

Figure 5: Local Run Manager User Interface—With Local Run Manager, runs can be set up, organized, and analyzed directly on the sequencing instrument.

Simplified Bioinformatics

Data analysis with the MiniSeq System requires no informatics expertise or command-line experience. The MiniSeq System features Local Run Manager software, an on-instrument system for creating a run, monitoring status, and analyzing sequencing data (Figure 5). With Local Run Manager, on-instrument data analysis can be automatically performed upon completion of the sequencing run. The data analysis modules generate simple reports for a wide range of sequencing applications. The modular design allows users to install and update individual analysis modules as needed.

In addition, sequencing data generated with the MiniSeq System can be instantly transferred, stored, and analyzed in the BaseSpace computing environment (cloud-based or onsite). BaseSpace Targeted Resequencing Software Apps provide expert-preferred data analysis tools packaged in an intuitive, click-and-go user interface designed for informatics novices (Figure 5). These Apps deliver optimized pipelines that support a range of common sequencing data analysis needs such as alignment, variant calling, and more. For enrichment workflows, the BaseSpace Isaac™ Enrichment App⁵ aligns targeted sequence reads with the ultrafast Isaac Aligner⁶ and performs variant calling with the Starling Variant Caller.⁶ For amplicon workflows, the TruSeq Amplicon App⁷ performs a banded Smith-Waterman alignment and enables

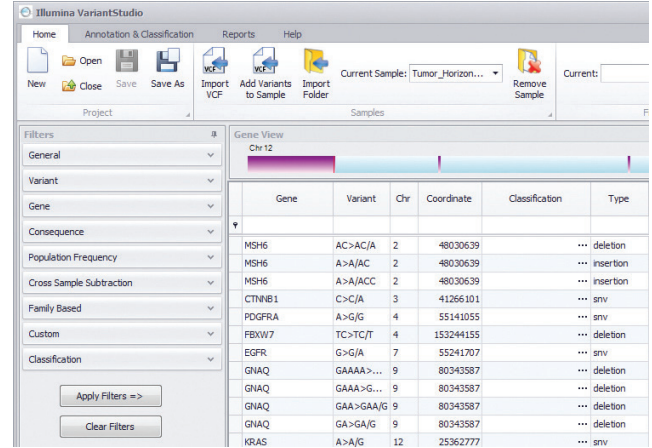


Figure 6: VariantStudio User Interface—Quickly identify, classify, and report disease-relevant variants with Illumina VariantStudio annotation software.

variant calling with the genome analysis toolkit (GATK 1.6),⁸ Isaac Variant Caller,⁶ or the Illumina-developed Somatic Variant Caller.⁹

For downstream analysis, the Illumina VariantStudio analysis software enables identification and classification of disease-relevant variants as well as generation of structured, detailed reports (Figure 6). Additionally, BaseSpace Apps generate output files that can be directly input into a broad range of data analysis tools. The BaseSpace Environment includes a growing community of developers who use and provide software tools for visualization, analysis, and sharing. This NGS ecosystem provides one of the largest collections of commercial and open-source analysis tools currently available.

NGS Targeted Resequencing vs Traditional Technologies

While traditional methods, such as CE-based sequencing and PCR can be used to interrogate specific regions of interest, NGS targeted resequencing provides the most cost-effective approach to sequencing the broadest regions of interest with the highest sensitivity (Table 2).

Table 2: Comparison of CE Sequencing, q/RT-PCR, and NGS Targeted Resequencing

	CE Sequencing	q/RT-PCR	Targeted Resequencing
Benefits	<ul style="list-style-type: none"> Cost-effective sequencing for small stretches^a of DNA sequence Quick and simple workflow Current gold standard in sequencing 	<ul style="list-style-type: none"> High sensitivity^b Quick and simple workflow Capital equipment already placed in most labs 	<ul style="list-style-type: none"> Higher sequencing depth enables higher sensitivity (down to 1%)^b Higher discovery power (screen hundreds of genes simultaneously) Higher mutation resolution (nucleotide identity can be determined) Produce more data with the same amount of input DNA^d Higher sample throughput with sample multiplexing
Challenges	<ul style="list-style-type: none"> Low sensitivity (down to 20%)^b Low discovery power Not as cost-effective for large stretches^c of DNA sequence Low scalability due to increasing sample input requirements 	<ul style="list-style-type: none"> Can only interrogate a limited set of mutations Virtually no discovery power Limited mutation resolution Low scalability due to increasing sample input requirements 	<ul style="list-style-type: none"> Not as cost-effective for sequencing small stretches^a of DNA sequence Not as time-effective for sequencing small stretches^a of DNA sequence

a. small stretches = less than ~15-20 amplicons

b. sensitivity = allele frequency limit of detection

c. large stretches = more than ~15-20 amplicons

d. 10 ng DNA will produce ~1 kb with CE sequencing or ~300 kb with targeted resequencing (250 bp amplicon length × 1536 amplicons with TruSeq Custom Amplicon workflow)

